## (*E,E*)-10-(1,3-Dihydro-4,6-dihydroxy-7-methyl-3-oxoisobenzofuran-5-yl)-4,8-dimethyldeca-4,8-dienoic Acid: Total Synthesis and Role in Mycophenolic Acid Biosynthesis

By LINO COLOMBO, CESARE GENNARI, DONATELLA POTENZA, and CARLO SCOLASTICO\*

(Istituto di Chimica Organica dell'Università, Centro delle Sostanze Organiche Naturali del C.N.R., via Saldini 50,

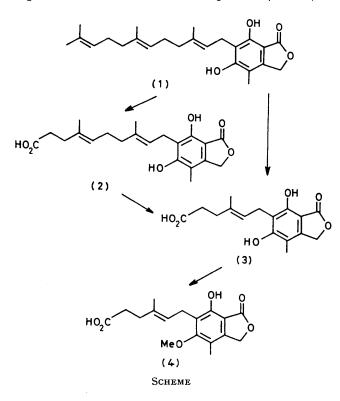
Milano, Italy)

and FABRIZIO ARAGOZZINI

(Cattedra di Microbiologia Industriale della Facoltà di Agraria dell'Università, via Celoria 2, Milano, Italy)

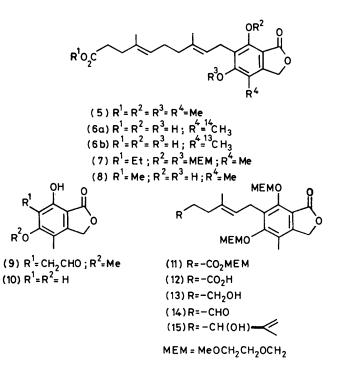
Summary The title compound (2), a prenylogue of mycophenolic acid (4), has been synthesized by two different routes; experiments show unambiguously that (2) is a biosynthetic intermediate of mycophenolic acid.

6-FARNESYL-5,7-DIHYDROXY-4-METHYLPHTHALIDE (1) is converted into mycophenolic acid (4) not only by direct oxidation of the central double bond,<sup>1</sup> but also by a two-stage, 'terminal-central, demolition process' (Scheme).



Previously reported radio g.l.c.-mass spectral data<sup>2</sup> are consistent with the latter process but alone do not prove the intermediacy of the title compound (2) in mycophenolic acid biosynthesis. Proof is given in this paper by isolating (2) and studying the incorporation of labelled (2) into mycophenolic acid. A synthetic sample of (2) was methylated  $(CH_2N_2; MeOH)$  to give (5). Comparison (g.l.c.mass spectroscopy) between the synthetic compound (5) and the fermentation compound, obtained by methylation  $(CH_2N_2; MeOH)$  of the mycelium extracts and silica gel chromatographic purifications,<sup>†</sup> shows that (2) is a natural metabolite of *Penicillium brevicompactum* [g.l.c. conditions: 1% SE 30, 220 °C; m/e 416 (10·4), 385 (6·9), 384 (9·2), 275 (43·0), 221 (100), 207 (31·4), 195 (53·0), and 141 (44·0%)].

Administration of the <sup>14</sup>C-labelled acid (**6a**) to the *in vivo* culture led to the formation of labelled mycophenolic acid (total incorporation 22%).<sup>‡</sup> Introduction of the <sup>13</sup>C-labelled acid (**6b**) yielded 28% molar incorporation into mycophenolic acid after the same fermentation time (3 days). These data show that (**2**) is a biosynthetic intermediate of mycophenolic acid. The whole oxidation sequence, in the conversion of (**1**) into (**4**), is currently under investigation, in particular the relative importance of the direct-central and terminal-central pathways.

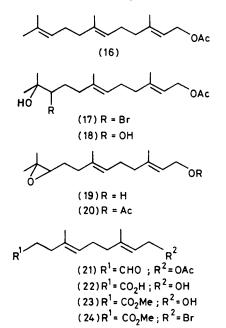


† Ca. 10 mg of (5) were obtained from the mycelium extracts derived from ten flasks of culture (L. Canonica, W. Kroszczynski, B. M. Ranzi, B. Rindone, E. Santaniello, and C. Scolastico, J.C.S. Perkin I, 1972, 2639).

<sup>‡</sup> The incorporation specificity was tested by ozonolysis of (4) to the aldehyde (9) (L. Canonica, W. Kroszczynski, B. M. Ranzi, B. Rindone, E. Santaniello, and C. Scolastico, *J.C.S. Perkin I*, 1972, 2639): the molar activity value did not change.

Two different approaches were used for the synthesis of the prenylogous acid (2). In the semisynthetic route, mycophenolic acid (4) was converted into the protected aldehyde (14) by the following sequence: demethylation to (3) (LiI; collidine; 70% yield),<sup>3</sup>  $\beta$ -methoxyethoxymethylation to (11) (MEMCl; Et<sub>3</sub>N; CH<sub>2</sub>Cl<sub>2</sub>-THF 5:1; 100% yield),<sup>4</sup> hydrolysis to (12) (NaOH; H<sub>2</sub>O-THF; 100% yield), reduction to (13) (ClCO<sub>2</sub>Me; Et<sub>3</sub>N; THF-NaBH<sub>4</sub>; H<sub>2</sub>O; 90% yield),<sup>5</sup> and oxidation of the alcohol (pyridinium chlorochromate;  $CH_2Cl_2$ ; AcONa; 80% yield) (THF = tetrahydrofuran).<sup>6</sup> Treatment of (14) with prop-2-enyllithium (Et<sub>2</sub>O-THF 5:1; -60 °C) gave the alcohol (15) (50% yield after Florisil chromatography) which underwent a Claisen-type transposition<sup>7</sup> [MeC(OEt)<sub>3</sub>; EtCO<sub>2</sub>H; 130 °C] to form the compound (7) in 40% yield. Hydrolysis of the MEM ethers (HCl; H<sub>2</sub>O-THF; 50% yield) and of the ethyl ester (NaOH, H<sub>2</sub>O; 100% yield) of (7) gave (E,E)-10-(1,3-dihydro-4,6-dihydroxy-7-methyl-3-oxoisobenzofuran-5-yl)-4,8-dimethyldeca-4,8-dienoic acid (2).

In the total synthesis, (E,E)-farmesol was converted into the terminal diol (18) by the following sequence: acetylation to (16) (Ac<sub>2</sub>O; pyridine), oxidation to the bromohydrin (17) (N-bromosuccinimide; Bu<sup>t</sup>OH-H<sub>2</sub>O),<sup>8</sup> alkaline treatment to (19) (K<sub>2</sub>CO<sub>3</sub>; MeOH-H<sub>2</sub>O), acetylation to (20) (Ac<sub>2</sub>O; pyridine), and acid hydrolysis of the epoxide (HClO<sub>4</sub>; diglyme-H<sub>2</sub>O). Oxidation of the diol (18) gave the aldehyde (21) (NaIO<sub>4</sub>;  $H_2O-THF$ ), which was in turn oxidised to form the acid (22) (Ag<sub>2</sub>O; NaOH; dioxan-H<sub>2</sub>O). Methylation of (22) gave the ester (23) (CH<sub>2</sub>N<sub>2</sub>; Et<sub>2</sub>O) which was brominated (CBr<sub>4</sub>; PPh<sub>3</sub>; MeCN)<sup>9</sup> to give (24). The reaction of the allylic bromide (24) with the diphenol (10) and Ag<sub>2</sub>O in dioxan-water (96:4)<sup>10</sup> gave the C-alkylated product (8) in 20% yield. Final hydrolysis (NaOH, H<sub>2</sub>O) gave the prenylogous acid (2).



The second synthetic route is more convenient, especially for the introduction of the label. The same reactions, using labelled (10), § gave the labelled acids (6).

Interest in the prenylogous acid (2) is not only limited to biosynthetic studies. Its pharmacological activity could be interesting, because of the structural analogy between (2), mycophenolic acid (4), and other anti-psoriasis agents.<sup>11</sup>

We thank Eli-Lilly Internat. Corp. for kindly giving us the mycophenolic acid.

(Received, 3rd July 1979; Com. 715.)

[Me-14C] and [Me-13C]-phthalides (10) were obtained by a modification of the literature procedure (W. R. Logan and G. T. Newbold, J. Chem. Soc., 1957, 1946) using labelled formaldehyde in the chloromethylation step.

<sup>1</sup>L. Colombo, C. Gennari, and C. Scolastico, J.C.S. Chem. Comm., 1978, 434. <sup>2</sup>C. P. Nulton and I. M. Campbell, Canad. J. Microbiol., 1978, 24, 199; C. P. Nulton, J. D. Naworal, I. M. Campbell, and (in part) E. W. Grotzinger, Analyt. Biochem., 1976, 75, 219.

<sup>3</sup> I. T. Harrison, Chem. Comm., 1969, 616.

<sup>4</sup> E. J. Corey, J-L. Gras, and P. Ulrich, Tetrahedron Letters, 1976, 809.
<sup>5</sup> D. F. Jones and S. D. Mills, J. Medicin. Chem., 1971, 14, 305.
<sup>6</sup> E. J. Corey and J. W. Suggs, Tetrahedron Letters, 1975, 2647.
<sup>7</sup> W. S. Johnson, L. Werthemann, W. R. Bartlett, T. J. Brocksom, T. Li, D. J. Faulkner, and M. R. Petersen, J. Amer. Chem. Soc., 1970, 92, 741.
<sup>6</sup> F. F. van Tamplon, Accounts Chem. Res. 1968, 1, 111, and references therein.

<sup>8</sup> E. E. van Tamelen, Accounts Chem. Res., 1968, 1, 111, and references therein.

E. H. Axelrod, G. M. Milne, and E. E. van Tamelen, J. Amer. Chem. Soc., 1970, 92, 2139.
 L. Canonica, B. Rindone, E. Santaniello, and C. Scolastico, Tetrahedron, 1972, 28, 4395.

<sup>11</sup> Drugs of the Future, 1978, Vol. III, No. 8, p. 594, and references therein; Unlisted Drugs, Nov. 1977, 29: 176 o, and references therein.